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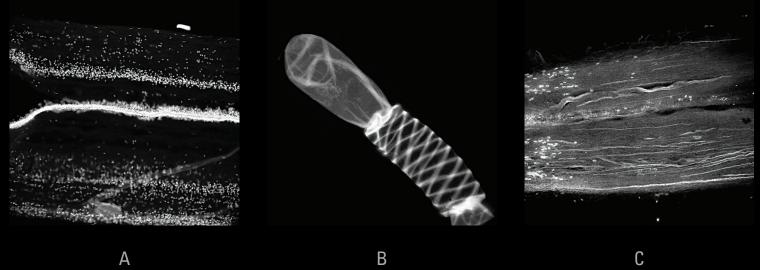
> Cat. No. SCAS0136E02 DEC/2018 HPK Created in the USA





COLUMN SALAR

The ORCA-Fusion, built from the sensor up, balances the complex nuances of camera features to provide beautiful images and robust data at all lights levels, but especially in tough low-light conditions. The exceptionally low and highly uniform read noise of



the ORCA-Fusion means that when the sample emits even just a handful of photons, either by default or by experimental design, they are not lost in the noise, but detected and reliably quantified. After all, when you want to hear a whisper it's best to be in a quiet place.

ORCA-FUSION SPECIFICATIONS

Camera Readout noise (electrons, rms) **ORCA-Fusion** Fast scan Standard scan Ultra quiet scan Product Number C14440-20UP Quantum efficiency @ 400 nm @ 550 nm Pixel Size @ 700 nm @ 800 nm 6.5 µm x 6.5 µm Full well capacity Effective number of pixels Dynamic range ^{*} 2304 x 2304 **Conversion factor Cooling Temperature Effective Area** With forced-air 14.976 mm x 14.976 mm Water cooled Dark current (electrons/pixel/sec @ -5 °C @ -15 °C **Dark offset** FUSION IMAGES Dark signal non-uniformity (DSN A. Image of a regenerated lamprey spinal cord labeled Photo response non-uniformity (P @7500 electrons with DAPI, which reveals the Linearity error *1 (EMVA 1288 stan distribution of cells throughout the spinal cord. The brightest **Readout modes** signal in the center of the spinal cord is the central canal. **Readout times at full resolution** Fast scan Credits: E. Guadarrama and Standard scan J. Morgan (Marine Biological Ultra quiet scan Laboratory). Lightsheet readout (fast scan) B. Plant tissue culture induced Row interval time to make xylem. Fluorescent Readout time at full resolution Readout modes bands are cell wall thickenings Readout directions needed to reinforce the cell wall for water transport. **Exposure times** Credits: Sample prepared by Fast scan T. Baskin, UMass Amherst and Standard scan Ultra quiet scan provided by R. Oldenbourg, Marine Biological Laboratory **Trigger modes** Trigger delay function **C.** Image of a regenerated Trigger output lamprev spinal cord labeled Input trigger connector with a neurofilament antibody Output trigger connectors Master pulse mode which reveals numerous regenerating axons. Credits: **Digital output** E. Guadarrama and J. Morgan Interface (Marine Biological Laboratory). Lens mount Software

*1 Typical value

1	1.4 1.0 0.7
	65 % 80 % 70 % 50 %
	15,000 electrons 21 400:1 ^{*2} 0.24 electrons / count
	-5 °C (Ambient temperature: +25 °C) *3 -5 °C (Water temperature: +25 °C) *3
econd) *1	0.5 0.2 100 counts
NU)	0.3 electrons rms
(PRNU) ndard)	0.06 % rms 0.5 %
	Full resolution, digital binning (2 x 2, 4 x 4), sub-array, lightsheet
*4	11.22 ms (89 fps with CoaXPress or 31.6 fps with USB 3.0) 42.99 ms (23.2 fps with CoaXPress or USB 3.0) 184.4 ms (5.4 fps with CoaXPress or USB 3.0)
	4.868 μs to 963.8 μs ^{*4} 11.22 ms to 2.221 s ^{*4} Full resolution, sub-array Top to bottom readout / Bottom to top readout
	17 μs to 10 s (4.87 μs step) 65 μs to 10 s (18.65 μs step) 280 μs to 10 s (80.00 μs step)
	Edge, Level, Sync readout, Start, Global reset edge, Global reset level, programmable Yes Global exposure timing, trigger ready, low, high SMA x1 SMA x 3 Free running / start trigger / burst
	16 bit ^{*5} / 12 bit / 8 bit CoaXPress (6.25 Gbpsx2 lane) and USB 3.0 Super Speed ^{*6} C-mount (Standard) / F-mount (Part Number TBD) HCImage, LabVIEW, MATLAB, μManager